

# blood

2005 105: 4845-4848  
Prepublished online Mar 1, 2005;  
doi:10.1182/blood-2004-12-4700

## Phenotypic characterization of the human myeloma cell growth fraction

Nelly Robillard, Catherine Pellat-Deceunynck and Régis Bataille

---

Updated information and services can be found at:

<http://bloodjournal.hematologylibrary.org/cgi/content/full/105/12/4845>

Articles on similar topics may be found in the following *Blood* collections:

[Neoplasia](#) (4216 articles)

[Cell Cycle](#) (231 articles)

[Brief Reports](#) (1184 articles)

---

Information about reproducing this article in parts or in its entirety may be found online at:

[http://bloodjournal.hematologylibrary.org/misc/rights.dtl#repub\\_requests](http://bloodjournal.hematologylibrary.org/misc/rights.dtl#repub_requests)

Information about ordering reprints may be found online at:

<http://bloodjournal.hematologylibrary.org/misc/rights.dtl#reprints>

Information about subscriptions and ASH membership may be found online at:

<http://bloodjournal.hematologylibrary.org/subscriptions/index.dtl>

Blood (print ISSN 0006-4971, online ISSN 1528-0020), is published semimonthly by the American Society of Hematology, 1900 M St, NW, Suite 200, Washington DC 20036.

Copyright 2007 by The American Society of Hematology; all rights reserved.



## Brief report

## Phenotypic characterization of the human myeloma cell growth fraction

Nelly Robillard, Catherine Pellat-Deceunynck, and Régis Bataille

In this study we quantified the proliferation rate of normal and malignant plasma cells (PCs) by *ex vivo* incorporation of 5-bromo-2'-deoxyuridine (BrdU; labeling index, LI) using flow cytometry. We show that all bone marrow PCs, either normal or malignant, include a subset of proliferating PCs present within the CD45<sup>bright</sup> fraction. Indeed, medullary normal and malignant PCs were always heterogeneous for CD45 expression, and prolifera-

tion was always restricted primarily to the CD45<sup>bright</sup> compartment. Moreover, an inverse correlation was found between LI or CD45 and B-cell lymphoma 2 (Bcl-2) in both malignant and normal PCs, the most proliferating CD45<sup>bright</sup> PCs have the lowest Bcl-2 expression. We investigated expression of molecules of interest in multiple myeloma (MM)—that is, CD138, CD19, CD20, CD27, CD28, CD56, and CD11a—to further characterize the CD45<sup>bright</sup> frac-

tion. Among all of these molecules, only CD11a was exclusively expressed by CD45<sup>bright</sup> proliferating myeloma cells. In conclusion, proliferating myeloma cells are characterized by the specific CD45<sup>bright</sup> CD11a<sup>pos</sup> Bcl-2<sup>low</sup> phenotype. (Blood. 2005;105:4845-4848)

© 2005 by The American Society of Hematology

## Introduction

Multiple myeloma (MM) is primarily conceptualized as an accumulative disease. Indeed, most studies evaluating the labeling index (LI) of plasma cells (PCs) revealed that it rarely exceeded 1%. However, as early as 1981, Drewinko et al<sup>1</sup> showed that *in vivo* myeloma cells were generally nonproliferating, although a small fraction of them could proliferate (growth fraction, GF). Thus, they proposed the concept that this minor GF could give rise to the major nonproliferating fraction.

Myeloma cells either lack or express a weak to intermediate level of CD45.<sup>2,3</sup> However, in previous studies we have shown that CD45 and also CD11a are frequently expressed by a subset of myeloma cells only.<sup>3</sup> It has been demonstrated that CD45 expression is highly correlated with the proliferation rate of myeloma cells.<sup>4,5</sup> With regard to normal counterparts, PCs are heterogeneous in terms of CD45 phenotype. Recently, Medina et al<sup>6</sup> confirmed the association of maturity with decreasing CD45 expression.<sup>7-9</sup> Generation of PCs from B cells, mainly studied *in vitro* in humans, is a multistep process that involves both proliferation and maturation/differentiation.<sup>10-13</sup> The aim of this study was to evaluate the proliferation of different types of normal PCs in relation to their phenotype, especially to CD45, to understand the biology of polyclonal and monoclonal PC expansions that are reactive plasmacytoses (RP) and MM, respectively.

patients with reactive plasmacytosis, and tonsil samples were obtained and prepared as described.<sup>3,13</sup> Approval was obtained from the Nantes University Hospital Institutional Review Board for these studies. Informed consent was provided according to the Declaration of Helsinki. Antibodies directed against (1) CD11a, CD19, CD45, CD138, Apo2.7 and (2) CD28, CD38, CD56, 5-bromo-2'-deoxyuridine (BrdU) were from Beckman Coulter (Miami, FL) and BD Biosciences (San Jose, CA), respectively.

## Cell staining

Mononuclear cells (MNCs;  $0.5-3 \times 10^6$ ) were stained in a 4-color assay with anti-CD45–fluorescein isothiocyanate (FITC; J33), anti-CD138–phycoerythrin cyanine 5 (PECy5; B-B4), anti-CD38–APC (HB7) and control-PE, anti-CD56–PE (MY31), anti-CD28–allophycocyanin (PE; L293), anti-CD19–PE (J4.119), anti-APO2.7–PE (2.7A6A3), anti-CD11a–PE (25.3) monoclonal antibodies (mAbs) as described.<sup>14</sup> To determine CD45-FITC fluorescent staining, MNCs were stained with isotype-FITC, anti-CD38–APC, anti-CD45–PECy5, and anti-CD138–PE mAbs. For intracellular Bcl-2 staining, MNCs were first stained with anti-CD38–APC, anti-CD45–PECy5, and anti-CD138–PE mAbs, then with anti-Bcl-2–FITC (124; Dako, Glostrup, Denmark) or control isotype-FITC mAbs after permeabilization (Intra Prep; Beckman Coulter). For LI, cells were incubated overnight with or without 50  $\mu$ M BrdU (5-bromo-2'-deoxyuridine; Sigma, St Louis, MO) at 37°C in culture medium, then stained with anti-CD45–APC, anti-CD38–PE, and anti-CD138–PECy5 mAbs (with anti-CD45–APC and anti-CD38–PE mAbs for tonsil and blood PCs), permeabilized, and stained with anti-BrdU as described.<sup>12</sup>

## Study design

## Samples and reagents

Forty-nine consecutive patients with MM (25 at diagnosis, 24 at relapse) were included in this study. Bone marrow and blood samples from healthy donors or

## Flow cytometry analysis

Data were acquired by means of a Becton Dickinson fluorescence-activated cell sorting FACSCalibur with CellQuest Pro software (BD Biosciences).<sup>14</sup> PCs were identified using a sequential gating strategy (Figure 1).

From the Institut National de la Santé et de la Recherche Médicale (INSERM) UMR601, Département de Recherche en Cancérologie, Nantes, France; and the Central Laboratory of Hematology University Hospital, Nantes, France.

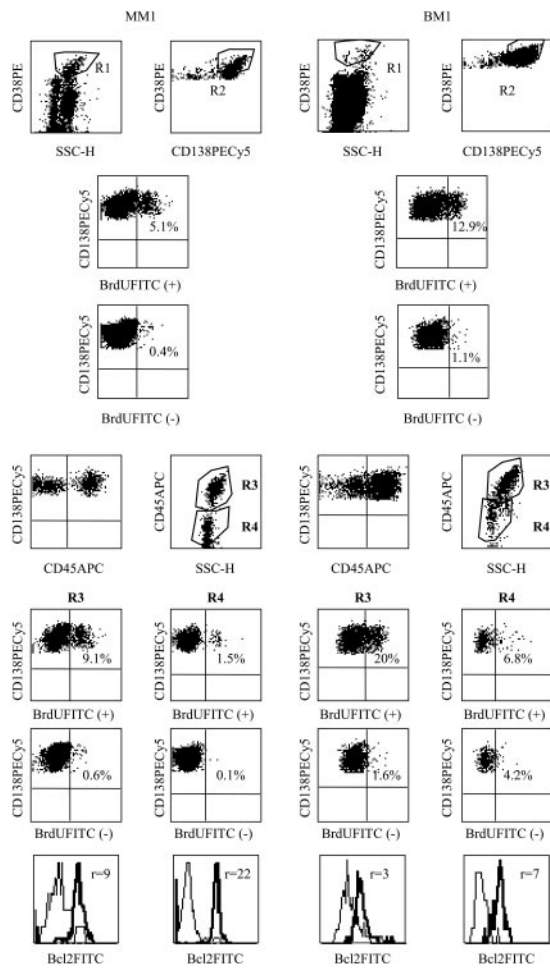
Submitted December 9, 2004; accepted February 10, 2005. Prepublished online as *Blood* First Edition Paper, March 1, 2005; DOI 10.1182/blood-2004-12-4700.

Supported by the Ligue Nationale Contre le Cancer (Equipe labellisée 2004).

**Reprints:** Catherine Pellat-Deceunynck, INSERM UMR601, Département de Recherche en Cancérologie, 9 quai Moncoussu, 44093 Nantes cedex 01, France; e-mail: cpellat@nantes.inserm.fr.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 U.S.C. section 1734.

© 2005 by The American Society of Hematology



**Figure 1. CD45 phenotype and LI of myeloma and bone marrow PCs.** PC phenotype analysis was performed in 2 steps. In the first step, 15 000 total events were acquired to draw a PC gate in the side scatter (SSC)/CD38<sup>++</sup> dot-plot (R1). In the second step, an acquisition of 15 000 PCs (at least 1000 PCs for very low plasmacytoses) was performed through the R1 live-gate. PCs were identified by coexpression of CD38 and CD138 (gate R2). A third region was set on the light scatter of the cells satisfying both R1 and R2 to exclude debris or apoptotic PCs with a low CD138 expression and a characteristic light-scatter distribution.<sup>12</sup> The lack of Apo 2.7 staining in the gated population confirmed that cells were not apoptotic.<sup>15</sup> Then 2 PC subpopulations were identified on the CD45 versus SSC dot plot: R3 was set around PC with a large SSC expressing a high level of CD45, while R4 was set around PC with low SSC and negative or weak CD45 expression. The analyses of the phenotype and the LI were performed in these 2 PC subpopulations, separately and simultaneously. The percentage of BrdU<sup>+</sup> PCs within the population incubated with BrdU (BrdU<sup>+</sup>) or without (control, BrdU<sup>-</sup>) were indicated within the cytograms. For example, in patient MM1 the global LI was 1.7% (3.1%–1.4%) but 8.5% (9.1%–0.6%) in R3 and 1.4% (1.5%–0.1%) in R4. Overlay histograms represent the immunofluorescence of Bcl-2 (thick line) over the control (thin line). *r* indicates the ratio of MFI.

### Statistical analysis

Statistical analyses were performed using nonparametric Wilcoxon rank sum, Spearman correlation coefficient, and the sign tests.

## Results and discussion

### Myeloma cells and bone marrow PCs were heterogeneous: proliferation was restricted to the CD45<sup>bright</sup> SSC<sup>high</sup> compartment

The percentage of BrdU-positive myeloma cells was low, generally around 1% (*n* = 49 patients). In all patients, myeloma cells were heterogeneous for CD45 expression and SSC, allowing a delinea-

tion of 2 compartments: CD45<sup>bright</sup>SSC<sup>high</sup> and CD45<sup>int/low</sup>SSC<sup>low</sup>. The CD45<sup>bright</sup>SSC<sup>high</sup> compartment was always present, representing 12% (median) of myeloma cells (illustrated in Figure 1). The second compartment CD45<sup>int/low</sup>SSC<sup>low</sup> represented the majority of tumor cells (88%). We observed that the LI of the CD45<sup>bright</sup>SSC<sup>high</sup> compartment was always greater than that of the CD45<sup>int/low</sup>SSC<sup>low</sup> compartment and far above 1% (median, 6.8%; *n* = 49; Table 1). In general, the CD45<sup>bright</sup>SSC<sup>high</sup> compartment had a 6.8 higher proliferative level than the CD45<sup>int/low</sup>SSC<sup>low</sup> compartment (*P* < .01). As outlined in Table 1, the LI of the CD45<sup>bright</sup> compartment could reach 40% of myeloma cells, which was similar to what we observed in reactive plasmacytoses,<sup>12</sup> suggesting that in these patients, almost all CD45<sup>bright</sup> myeloma cells were proliferating. Of note, the total LI correlated highly with the LI of the CD45<sup>bright</sup> compartment (*r* = 0.81, *P* < .001) and less with that of the CD45<sup>low</sup> compartment (*r* = 0.44, *P* < .01).

As observed for myeloma cells, 2 compartments of PCs delineated by CD45 and SSC were found in normal bone marrows (*n* = 11, illustrated in Figure 1). The CD45<sup>bright</sup>SSC<sup>high</sup> compartment, which represented 65% of the PCs, was highly proliferative (LI = 18.4% for BM1). The second CD45<sup>low</sup>SSC<sup>low</sup> compartment included 35% of the PCs with a much lower LI: 2.6% (BM1 and Table 1). A subset of PCs which lacked CD45 expression was observed in 2 cases.

### Tonsillar and peripheral PCs were homogeneous and were capable of proliferation

In Figure 2, we show that PCs from both tonsils and blood, the prototypes of immature PCs, were homogeneously CD45<sup>bright</sup> and capable of proliferation (LI > 10%). Reactive PCs turned out to be similar to blood PCs with higher LI.

### BrdU<sup>+</sup> myeloma cells expressed lower Bcl-2 levels

Since we have previously observed that Bcl-2 inversely correlates with the LI in both reactive PCs and myeloma cells,<sup>16</sup> we looked for a correlation within myeloma and PC subsets (Figures 1-2). We found that CD45<sup>bright</sup> myeloma cells always expressed a lower level of Bcl-2 compared with that of CD45<sup>low/neg</sup> myeloma cells (*P* < .01). In normal bone marrow (BM), a significant increase in Bcl-2 MFIR (mean of fluorescence intensity ratio) was observed in the CD45<sup>low</sup> bone marrow PCs in comparison with that of the CD45<sup>bright</sup> PCs: 7.4 (4.9-13) versus 4.2 (3.1-9), *P* < .01. Concerning tonsillar and circulating PCs, we found a highly significant negative correlation between Bcl-2 MFIR and the LI (*r* = -0.854, *P* < .001). This inverse correlation in PCs is in agreement with data showing that (1) Bcl-2 negatively controls the proliferation of different human cells types,<sup>17</sup> and (2) Bcl-2 transgenic mice have increased B-cell memory and PC-producing immunoglobulins.<sup>18</sup> Up-regulation of Bcl-2 thus appears to be a critical event during this process of maturation with proliferation decrease and arrest.

### Phenotypic characterization of the CD45<sup>bright</sup> SSC<sup>high</sup> proliferating compartment in MM: only CD11a expression was restricted to the CD45<sup>bright</sup> SSC<sup>high</sup> subset

We further characterized the phenotype of the CD45<sup>bright</sup> myeloma cells using relevant MM markers (CD19, CD20, CD27, CD28, CD56). As illustrated in Figure 3, CD19, CD27, CD28, and CD56 were expressed or not by myeloma cells, and their expression was unrelated to CD45 subsets. However, we found that CD11a

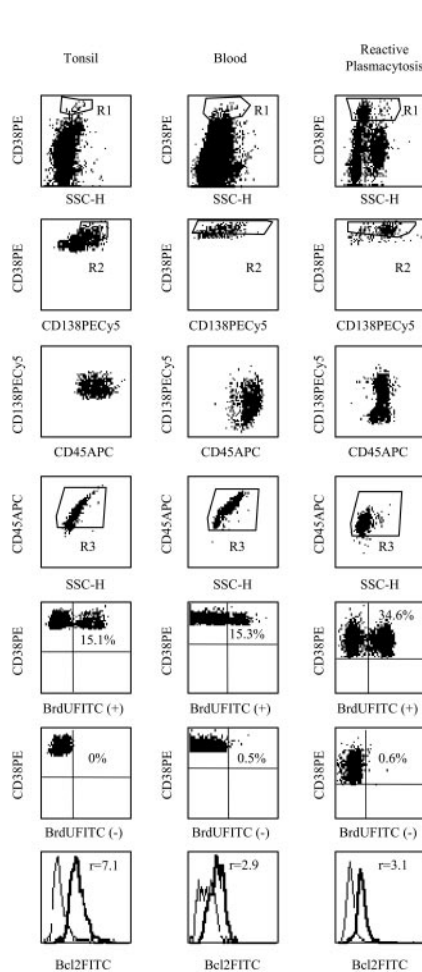
**Table 1. CD45 expression and labeling index of PCs**

Cell type	Cells, %		MFIR		PCLI, %	
	Median	Range	Median	Range	Median	Range
<b>Multiple myeloma</b>						
CD45 <sup>bright</sup> SSC <sup>high</sup>	12	2-68	34	10-92	6.8	0.5-40
CD45 <sup>low</sup> SSC <sup>low</sup>	88	32-98	1.2	1-3.8	1	0.2-16.6
<b>Normal bone marrow</b>						
CD45 <sup>bright</sup> SSC <sup>high</sup>	65	45-87	29.8	15-46.3	20.5	2.6-38
CD45 <sup>low</sup> SSC <sup>low</sup>	35	13-55	1.7	1-4.2	3.6	0-12
<b>Tonsil</b>						
CD45 <sup>bright</sup> SSC <sup>high</sup>	100	43-100	75	43-97	11.4	7-14
CD45 <sup>low</sup> SSC <sup>low</sup>	0	0-57	—	—	—	—
<b>Peripheral blood</b>						
CD45 <sup>bright</sup> SSC <sup>high</sup>	100	100	24	14-50	15.9	12.5-17
CD45 <sup>low</sup> SSC <sup>low</sup>	0	0	—	—	—	—
<b>Reactive plasmacytosis</b>						
CD45 <sup>bright</sup> SSC <sup>high</sup>	100	100	67	39-95	33	23-52
CD45 <sup>low</sup> SSC <sup>low</sup>	0	0	—	—	—	—

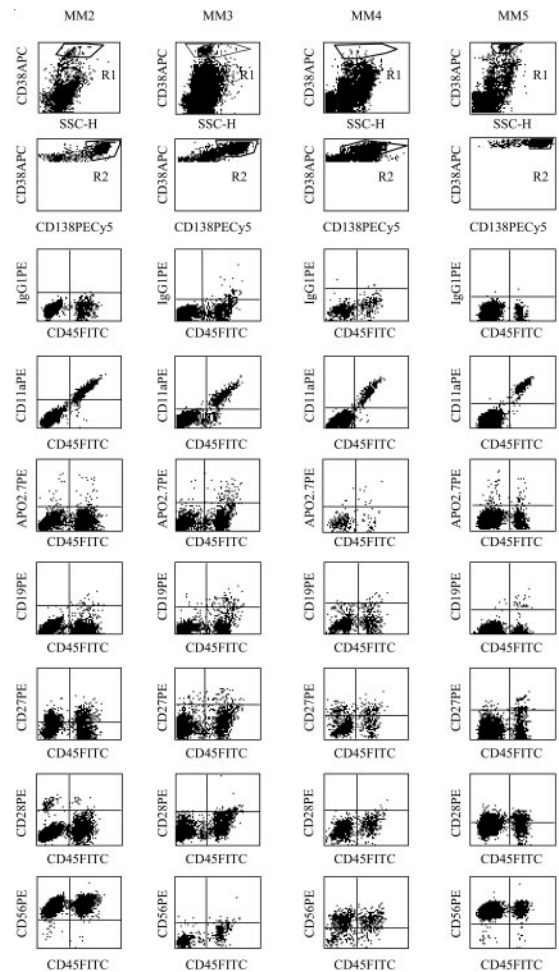
MFIR indicates mean of fluorescence intensity ratio; PCLI, plasma cell labeling index; and —, no data (no cells).

overlapped completely with the CD45<sup>bright</sup> compartment in all the patients studied (Figure 3), while CD45<sup>low/neg</sup> myeloma cells were negative for CD11a. Ahsmann et al<sup>19</sup> have published that lymphocyte function-associated antigen 1 (LFA-1) (CD11a-CD18) expression correlated with tumor growth in MM. LFA-1 is involved in

either homotypic or heterotypic interactions in MM. Indeed, human myeloma cells, like stromal cells, express the 3 ligands of LFA-1: intercellular adhesion molecule-1 (ICAM-1) (CD54), ICAM-2 (CD102), and ICAM-3 (CD50).<sup>3,20</sup> These interactions, restricted to



**Figure 2. CD45 phenotype and LI of tonsillar, peripheral normal, and reactive PCs.** Analysis of phenotype, LI, and Bcl-2 expression in PCs isolated from tonsil and blood. PCs were identified by CD38<sup>++</sup> expression (SSC/CD38) since CD138 expression was low in tonsil PCs and heterogeneous in peripheral PCs.



**Figure 3. Phenotype of CD45<sup>bright</sup> myeloma cells.** The phenotype of myeloma cells from 4 patients was determined in a 4-color assay. Myeloma cells were identified by coexpression of CD38 and CD138, and their phenotype (CD11a, CD19, CD27, CD28, CD56, Apo2.7) was analyzed in both CD45<sup>weak/neg</sup> and CD45<sup>bright</sup> subsets. Ig indicates immunoglobulin.

the most proliferative compartment, could have some important consequences for tumor behavior. For example, we have previously observed that ICAM-2/LFA-1 interactions were involved in the (negative) control of myeloma cell growth through CD40.<sup>20</sup>

Interleukin 6 (IL-6) has been shown to be a survival and growth factor for human myeloma cells.<sup>21,22</sup> More recently, it has been demonstrated that IL-6 preferentially stimulates CD45<sup>bright</sup> myeloma cells to proliferate<sup>23,24</sup> through activation of CD45-associated src kinase.<sup>24</sup> Of note, CD45<sup>bright</sup> myeloma cells express more IL-6 receptors.<sup>25</sup> IL-6 is also a survival and proliferative factor for nonmalignant PCs.<sup>12,13,26</sup> Taken altogether, these data

show that CD45<sup>bright</sup> myeloma cells, like their normal counterparts (CD45<sup>bright</sup> normal PCs), could be the main target of IL-6 to sustain survival and proliferation.

In conclusion, we have found that all patients with MM have a small proliferative compartment of myeloma cells characterized by a bright expression of CD45 and a specific expression of CD11a as well as a low Bcl-2 expression (sensitive to apoptosis). This CD45<sup>bright</sup>CD11a<sup>pos</sup> population of myeloma cells could constitute the growth fraction as defined in vivo by Drewinko et al<sup>1</sup> more than 20 years ago. Thus, this “to be killed” population could be targeted through CD45- or CD11a-targeted therapies.

## References

- Drewinko B, Alexanian R, Boyer H, Barlogie B, Rubinow SI. The growth fraction of human myeloma cells. *Blood*. 1981;57:333-338.
- Jackson N, Ling NR, Ball JJ, Bromidge E, Nathan PD, Franklin IM. An analysis of myeloma plasma cell phenotype using antibodies defined at the Illrd international workshop on human leukocyte differentiation antigens. *Clin Exp Immunol*. 1988;72:351-356.
- Pellat-Deceunynck C, Barillé S, Puthier D, et al. Adhesion molecules on human myeloma cells: significant changes in expression related to malignancy, tumor spreading, and immortalization. *Cancer Res*. 1995;55:3647-3653.
- Pope B, Brown R, Gibson J, Joshua D. The bone marrow plasma cell labeling index by flow cytometry. *Cytometry*. 1999;38:286-292.
- Fujii R, Ishikawa H, Mahmoud MS, Asaoku H, Kawano MM. MPC-1<sup>-</sup>CD49e<sup>-</sup> immature myeloma cells include CD45<sup>+</sup> subpopulations that can proliferate in response to IL-6 in human myelomas. *Br J Haematol*. 1999;105:131-140.
- Medina F, Segundo C, Campos-Caro A, Gonzalez-Garcia I, Brieva JA. The heterogeneity shown by human plasma cells from tonsil, blood, and bone marrow reveals graded stages of increasing maturity, but local profiles of adhesion molecule expression. *Blood*. 2002;99:2154-2161.
- Terstappen LW, Johnsen S, Segers-Nolten IM, Loken MR. Identification and characterization of plasma cells in normal human bone marrow by high-resolution flow cytometry. *Blood*. 1990;76:1739-1747.
- Harada Y, Kawano M, Huang N, et al. Identification of early plasma cells in peripheral blood and their clinical significance. *Br J Haematol*. 1996;92:184-191.
- Ocqueteau M, Orfao A, Almeida J, et al. Immunophenotypic characterization of plasma cells from monoclonal gammopathy of undetermined significance patients. *Am J Pathol*. 1998;152:1655-1665.
- Jelinek DF, Lipsky PE. The role of B cell proliferation in the generation of immunoglobulin-secreting cells in man. *J Immunol*. 1983;130:2597-2604.
- Arpin C, Banchereau J, Liu YJ. Memory B cells are biased towards terminal differentiation: a strategy that may prevent repertoire freezing. *J Exp Med*. 1997;186:931-940.
- Jego G, Robillard N, Puthier D, et al. Reactive plasmacytoses are expansions of plasmablasts retaining the capacity to differentiate into plasma cells. *Blood*. 1999;94:701-712.
- Jego G, Bataille R, Pellat-Deceunynck C. Interleukin-6 is a growth factor for nonmalignant human plasmablasts. *Blood*. 2001;97:1817-1822.
- Robillard N, Avet-Loiseau H, Garand R, et al. CD20 is associated with a small mature plasma cell morphology and t(11;14) in multiple myeloma. *Blood*. 2003;102:1070-1071.
- Zhang C, Ao Z, Seth A, Schlossman SF. A mitochondrial membrane protein defined by a novel monoclonal antibody is preferentially detected in apoptotic cells. *J Immunol*. 1996;157:3980-3987.
- Puthier D, Pellat-Deceunynck C, Barillé S, et al. Differential expression of Bcl-2 in human plasma cell disorders according to proliferation status and malignancy. *Leukemia*. 1999;13:289-294.
- Huang D, O'Reilly L, Strasser A, Cory S. The anti-apoptosis function of Bcl-2 can be genetically separated from its inhibitory effect on cell cycle entry. *EMBO J*. 1997;16:4628-4638.
- Strasser A, Whittingham S, Vaux DL, et al. Enforced Bcl-2 expression in B-lymphoid cells prolongs antibody responses and elicits autoimmune disease. *Proc Natl Acad Sci U S A*. 1991;88:8661-8665.
- Ahmann EJM, Lokhorst HM, Dekker AW, Bloem AC. Lymphocyte function-associated antigen-1 expression on plasma cells correlates with tumor growth in multiple myeloma. *Blood*. 1992;79:2068-2075.
- Pellat-Deceunynck C, Amiot M, Robillard N, Wijdenes J, Bataille R. CD11a-CD18 and CD102 interactions mediate myeloma cell growth arrest induced by CD40 stimulation. *Cancer Res*. 1996;56:1909-1916.
- Kawano M, Hirano T, Matsuda T, et al. Autocrine generation and requirement of BSF-2/IL-6 for human multiple myelomas. *Nature*. 1998;332:83-85.
- Klein B, Zhang X-G, Jourdan M, et al. Paracrine rather than autocrine regulation of myeloma-cell growth and differentiation by Interleukin-6. *Blood*. 1989;73:517-526.
- Mahmoud MS, Ishikawa H, Fujii R, Kawano M. Induction of CD45 expression and proliferation in U-266 myeloma cell line by interleukin-6. *Blood*. 1988;92:3887-3897.
- Ishikawa H, Tsuyama N, Abroun S, et al. Requirements of src family kinase activity associated with CD45 for myeloma cells proliferation by interleukin-6. *Blood*. 2002;99:2172-2176.
- Hata H, Xiao H, Petrucci MT, Woodliff J, Chang R, Epstein J. Interleukin-6 gene expression in multiple myeloma: a characteristic of immature tumor cells. *Blood*. 1993;81:3357-3364.
- Kawano MM, Mihara K, Huang N, Tsujimoto T, Kuramoto A. Differentiation of early plasma cells on bone marrow stromal cells requires interleukin-6 for escaping from apoptosis. *Blood*. 1995;85:487-494.