

blood

2008 111: 4930-4933
Prepublished online Feb 27, 2008;
doi:10.1182/blood-2008-01-117770

Structural and numerical variation of FLT3/ITD in pediatric AML

Soheil Meshinchi, Derek L. Stirewalt, Todd A. Alonzo, Titus J. Boggon, Robert B. Gerbing, Jennifer L. Rocnik, Beverly J. Lange, D. Gary Gilliland and Jerald P. Radich

Updated information and services can be found at:

<http://bloodjournal.hematologylibrary.org/cgi/content/full/111/10/4930>

Articles on similar topics may be found in the following *Blood* collections:

[Neoplasia](#) (4216 articles)

[Brief Reports](#) (1191 articles)

[Clinical Trials and Observations](#) (2641 articles)

Information about reproducing this article in parts or in its entirety may be found online at:

http://bloodjournal.hematologylibrary.org/misc/rights.dtl#repub_requests

Information about ordering reprints may be found online at:

<http://bloodjournal.hematologylibrary.org/misc/rights.dtl#reprints>

Information about subscriptions and ASH membership may be found online at:

<http://bloodjournal.hematologylibrary.org/subscriptions/index.dtl>



Brief report

Structural and numerical variation of FLT3/ITD in pediatric AML

Soheil Meshinchi,¹⁻³ Derek L. Stirewalt,^{1,4} Todd A. Alonzo,^{3,5} Titus J. Boggon,⁶ Robert B. Gerbing,³ Jennifer L. Rocnik,⁷ Beverly J. Lange,^{3,6} D. Gary Gilliland,⁷ and Jerald P. Radich^{1,4}

¹Clinical Research Division, Fred Hutchinson Cancer Research Center, and ²Department of Pediatrics, University of Washington Medical Center, Seattle;

³Children's Oncology Group, Arcadia, CA; ⁴Department of Medicine, University of Washington Medical Center, Seattle; ⁵University of Southern California Keck School of Medicine, Los Angeles; ⁶Department of Pharmacology, Yale University School of Medicine, New Haven, CT; and ⁷Division of Hematology/Oncology, Department of Pathology, Brigham and Women's Hospital; the Howard Hughes Medical Institute, Harvard Medical School, Boston, MA

FLT3 internal tandem duplication (FLT3/ITD) is a common somatic mutation in acute myeloid leukemia (AML) with significant variation in the position, length, and number of duplications of the FLT3 gene. We evaluated these physical characteristics in FLT3/ITD-positive patients who were treated on CCG-2941/2961 and correlated them with clinical outcome. Fifty-nine of 77 FLT3/ITD-positive patients (77%) had a single ITD, 16 (21%) had 2 ITDs, and

2 (3%) had 3 ITDs. The length of the duplicated region varied from 6 to 51 amino acids, and in all cases amino acid residues Y591-Y597 were duplicated. Structural analysis demonstrated that Y591-Y597 encodes the switch and zipper regions of the juxtamembrane domain of FLT3. In addition, 24 of 77 patients (31%) had duplication of the critical STAT5 docking sites Y589/591. Patients with longer ITDs had a worse relapse-free survival

(19% vs 51%, $P = .035$), while the presence of more than 1 ITD was not clinically significant. Physical characteristics including the length of FLT3/ITD may influence FLT3 activation state by altering its structure and may impact response to therapy. (Blood. 2008;111:4930-4933)

© 2008 by The American Society of Hematology

Introduction

FLT3 internal tandem duplication (FLT3/ITDs) is present in approximately 15% of pediatric and 25% of adult acute myeloid leukemia (AML) patients¹⁻⁵ and has been associated with inferior outcome.^{1-3,6,7} However, nearly one-fourth of FLT3/ITD-positive patients survive, suggesting biological and clinical variation among FLT3/ITD-positive patients. FLT3/ITD-positive patients may be risk stratified using mutant/wild-type allelic ratios (ITD-AR).^{3,8} Other physical characteristics of FLT3/ITD, including length and position of the tandem duplication within the juxtamembrane domain, have been linked to clinical and biologic variation, where a duplication of a single codon may alter biologic activity of the FLT3 gene.⁹⁻¹¹ We previously reported that in adults with AML, patients with ITDs of more than 40 nucleotides had a lower complete remission (CR; 35% vs 67%) and worse survival (13% vs 26%) compared with those with shorter ITDs.⁹ In this report we examine the variability in ITD length, number of ITDs, and the position of the duplication in FLT3 coding sequence and correlate these parameters with clinical outcome in pediatric AML.

Methods

Patients and mutation analysis

The study population has been detailed elsewhere.¹² In brief, available samples from 647 patients age 0 to 21 years, enrolled on CCG 2941 and CCG-2961, were tested for FLT3/ITD. FLT3/ITD screening, allelic ratio (ITD-AR) determination, and sequencing was performed as previously

described.^{6,8,13} Informed consents for biology studies were obtained in accordance with the Declaration of Helsinki at study enrollment. This study was approved by the Institutional Review Board of the Fred Hutchinson Cancer Research Center.

Crystal structure analysis

The FLT3 crystal structure¹⁴ was downloaded from the Protein Data Bank^{15,16} (accession code 1RJB), and the structure was analyzed in O.¹⁷

Statistical methods

Data were analyzed from CCG-2941 and CCG-2961 through April 2005 and June 2005, respectively. Statistical analysis of outcome differences and end points analyzed has been detailed previously.⁸

Results and discussion

Characteristics of the region of duplication in FLT3/ITD

Recent work has highlighted several functional moieties within exon 14 of the *FLT3* gene where specific amino acid (aa) residues are involved in receptor activation. As these functional motifs of FLT3 are variably involved in FLT3/ITD, we evaluated the variation of FLT3/ITD in a cohort of pediatric AML patients. FLT3/ITD was identified in 77 of 630 patients tested (12%) and their ITD allelic ratio (ITD-AR, ITD to wild-type ratio) determined as previously described.⁸ Fifty-nine patients (77%) had a single duplication. The remaining 18 patients (23%) had

Submitted January 8, 2008; accepted February 24, 2008. Prepublished online as *Blood* First Edition paper, February 27, 2008; DOI 10.1182/blood-2008-01-117770.

The publication costs of this article were defrayed in part by page charge

payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 USC section 1734.

© 2008 by The American Society of Hematology

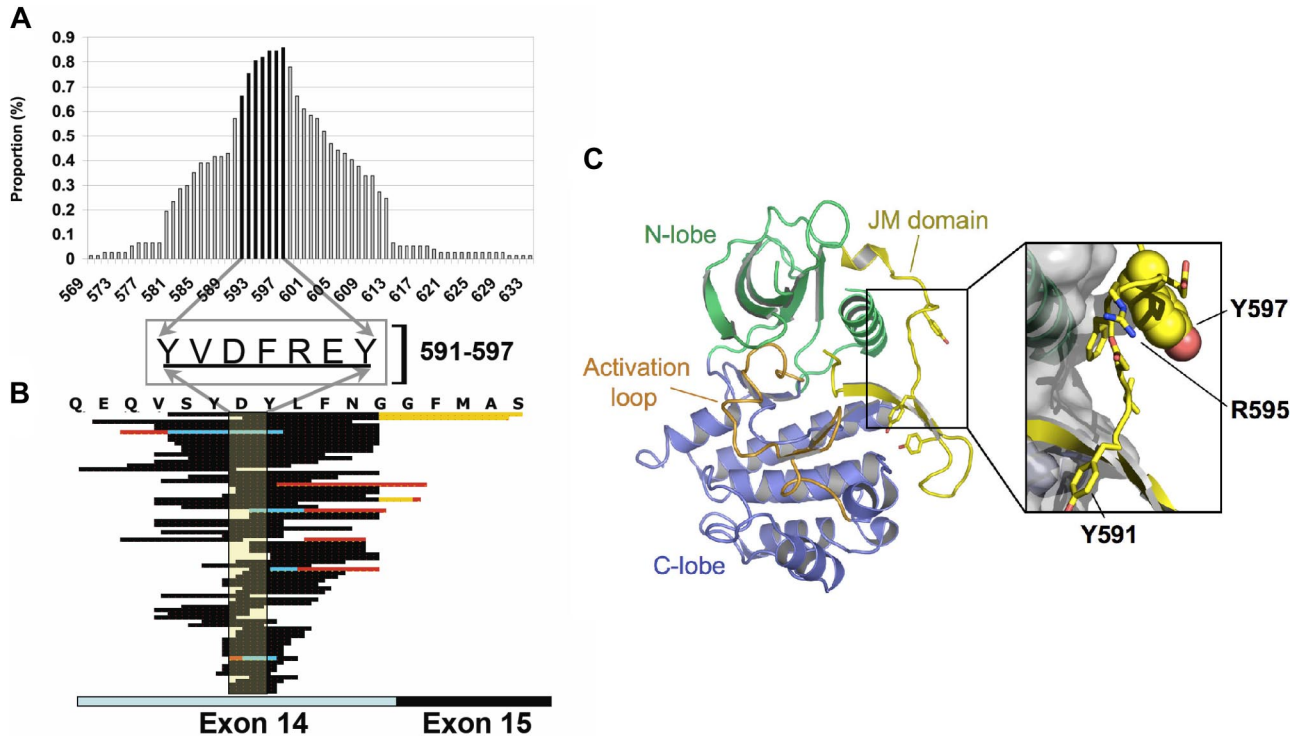


Figure 1. Region of duplication in FLT3/ITD. (A) Proportion of patients with involvement of specific aa residues in the duplicated region. Codons 591 to 597 are shown in darker bars, and specific aa residues in this region are shown in the center inset. (B) The region of duplication in all 77 patients are sorted by ITD length (longest to shortest) from top to bottom. The area of duplication of the major ITD peak is shown in dark highlighted area. In patients with additional ITDs, the duplicated region is shown in red with the area of overlap in blue. Codons 591 to 597 are shown in shaded box. (C) FLT3 structure showing the location of the JM domain and Y597. Ribbon representation of the crystal structure of FLT3 previously solved by Griffith et al.¹⁴ The kinase domain N-lobe is colored green and the C-lobe, blue. The activation loop is colored orange and the JM domain, yellow. Tyrosines 597, 591, and 589 are shown in stick format. The exploded view shows the molecular surface of the kinase domain and JM domain side-chain residues Y591 to Y597. Tyrosine 597 is depicted with space-filling spheres. This figure was made using the program Pymol (www.pymol.org).

either 2 ITDs ($N = 16$) or 3 ITDs ($N = 2$). All duplications involved exon 14 of the *FLT3* gene, and in 5 patients the duplication extended through intron 14 into exon 15. Forty of the 77 patients had a tandem duplication that was identical to native *FLT3* coding sequencing, whereas the remaining 37 patients had an insertion of 3 to 21 base pairs (median, 9 base pairs) preceding the duplication. The length of the ITDs ranged from 15 to 174 base pair (bp) with a median of 52 bp. The genomic localization of the duplication varied from patient to patient and overall spanned the entire length of exon 14 (Figure 1). Duplication of at least 1 of 7 aa residues from codon 591 to 597 was observed in all patients, and codon 597 was duplicated in 86% of the patients. In 5 cases, the duplication spanned intron 14 and involved exon 15 and codons 613 to 631. Vempati et al have reported similar results in a cohort of adult patients and have demonstrated that R595 is critical in ITD function.¹¹

Structural location of Y597

The crystal structure of autoinhibited FLT3 has revealed distinct regions within the JM domain, the JM binding motif (JM-B, Y572-M578), the JM switch motif (JM-S, V579-V592), and the JM zipper (JM-Z, D593-W603¹⁴; PDB code 1RJB). The amino acid residues that were observed in ITDs in all patients in this study (Y591-Y597) fall within the JM-S and JM-Z regions (Figure 1C). Tyrosine-597 is exposed on the surface of the protein and does not seem to be critical for maintenance of the autoinhibited conformation of FLT3 seen in the crystal structure. In this study, the residue most frequently observed within an ITD is Y597, however, other residues of the JM domain zipper (JM-Z) are also frequently duplicated. These duplications are strongly transforming and

result in constitutive activation of FLT3. The structural mechanisms of FLT3 activation by these duplications has not yet been elucidated, however, the crystal structure of autoinhibited FLT3 does shed some light on potential activating mechanisms. In this crystal structure the JM-Z region seems to play an important role in directing an optimal orientation of the autophosphorylation “switch” residues Y589 and Y591 and maintaining the autoinhibited conformation. JM-Z ITD insertions are expected to disrupt the autoinhibited conformation of the JM switch (JM-S) region, so preventing proper kinase inhibition by the JM binding (JM-B) region.^{11,14,18} Interruption of the optimal orientation of Y589 and Y591 is also expected to allow their constitutive phosphorylation. The reason for the prevalence of ITD mutations in the JM-Z region of the JM domain, compared with the JM-S and JM-B regions, has, however, not yet been elucidated.

Prognostic implication of FLT3/ITD length

We inquired whether ITD length may impact clinical outcome. We evaluated all ITD-length thresholds between the 10th and 90th quantiles for the ability to differentiate clinical outcome between those with shorter or longer than the particular ITD length. Cox regression analysis was used to compare the survival from diagnosis and relapse from CR for patients with higher or lower than a specific ITD-length threshold. ITD-length threshold of 48 bp had a hazard ratio (HR) of 2.3 for worse overall survival (OS) from study entry ($P = .013$) with no difference in CR rate (82% vs 81%, $P = .927$). Actuarial OS at 4 years from study entry for those with shorter ITD (≤ 48 bp) was 51% ($\pm 19\%$) compared with 17% ($\pm 15\%$) for those with ITD-length more than 48 bp ($P = .011$, Figure 2A). Actuarial OS at 4 years from CR for those with a shorter ITD was 67% ($\pm 21\%$) versus 21% ($\pm 19\%$) for those with

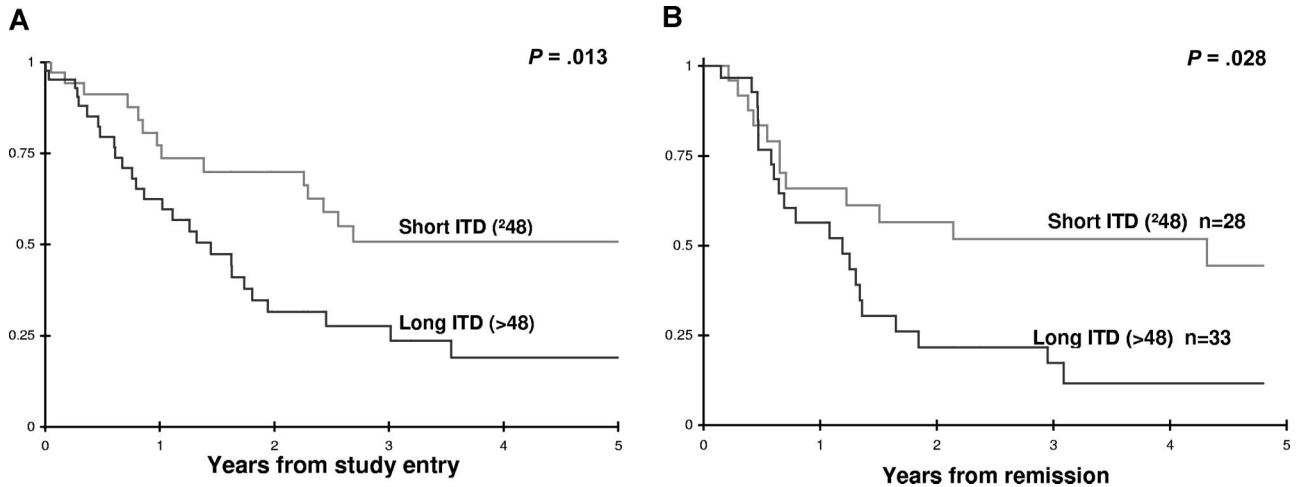


Figure 2. Clinical outcome for FLT3/ITD-positive patients with high versus low ITD length based on ITD length threshold of 48 bp. (A) Overall survival from study entry. (B) Relapse-free survival from complete remission.

longer ITDs ($P = .006$). Relapse-free survival at 4 years from CR for those with shorter and longer ITDs was 51% ($\pm 23\%$) and 19% ($\pm 16\%$), respectively ($P = .035$, Figure 2B). Corresponding OS and relapse-free survival (RFS) for patients without FLT3/ITD was 54% and 57%, respectively, similar to those with shorter ITDs. Thus, in FLT3/ITD-positive patients, those with shorter ITDs have a lower relapse rate and more favorable outcome than those with longer ITDs. Although we have demonstrated the significance for ITD length in adult⁹ and pediatric AML, other studies have not demonstrated such outcome differences based on ITD length,^{19,20} suggesting that additional factors (eg, nature of duplicated moieties) may modify the significance of ITD length.

Clinical implications of duplication of Y589-591

Recent studies have identified 2 candidate STAT5 docking sites within the FLT3 juxtamembrane domain coding region that can be disrupted by ITD. Tyrosine to phenylalanine substitution of residues 589 and 591 in the context of the FLT3-ITD maintains tyrosine kinase activity but abrogates STAT5 activation and fails to induce a myeloproliferative process in mice.¹⁰ We tested whether involvement of 589 to 591 codons in ITDs correlated with clinical outcome.

In 53 of 77 patients (69%) the ITDs involved the codons 589 to 591. Response to therapy was evaluated in those with and without 589 to 591 duplication. The remission induction rate for those with or without Y589/591 duplication was similar (88% vs 93%, $P = .6$). Relative risk of induction failure or post-remission relapse was 80% for those with Y589 to 591 duplication compared with 54% for those without ($P = .7$). The corresponding RFS at 5 years from CR was 26% and 46% for those with or without Y589/591 involvement, respectively ($P = .42$). There was no statistically significant difference in outcome for those with or without Y589/591 duplication, and any perceived difference needs to be evaluated in a larger study population.

Prognostic significance of presence of multiple ITDs

There have been suggestions that patients with multiple ITDs have a worse outcome than those with a single ITD.² We evaluated whether those with multiple ITDs have a different outcome compared with those with a single ITD. Fifty-nine of the 77 patients (77%) had a single duplication, and the remaining 18 patients (23%) had either 2 ITDs ($N = 16$) or 3 ITDs ($N = 2$). Overall survival at 5 years from study entry for

those with 1 ITD versus more than 1 ITD was 30% and 31% ($P = .52$), with a corresponding relapse rate at 5 years from remission of 70% and 69% ($P = .35$), demonstrating that the presence of multiple ITDs does not carry prognostic significance in this clinical context.

This study suggests that the length of the ITD may be important in disease prognosis. Whether it is the length of the ITD or the involvement of specific residues need yet to be clarified. It is possible that longer ITDs may involve multiple functional domains and more effectively disrupt the autoinhibitory regulation of the kinase activity. It is interesting to note that the lengths of the 3 JM regions, JM-B, JM-S, and JM-Z, are 7, 14, and 11 amino acids in length; a JM domain ITD length greater than 14 amino acids will necessarily duplicate residues from more than one of these regions, disrupting more than one of the structural requirements for autoinhibition. Thus, it is feasible that simultaneous disruption of more than one of the JM domain regions could underlie the prognostic significance of ITD length. Further studies are needed to elucidate the functional dependencies of the JM domain regions on one another. We also show that STAT5 binding sites are duplicated in one-third of the cases and may impact outcome, although the latter does not meet statistical significance. Collectively, these findings indicated that among the physical parameters that delineate between various FLT3-ITDs, increased length of the ITD is a poor prognostic indicator and may warrant alternative approaches to therapy in this cohort of pediatric patients.

Acknowledgments

This work was supported by National Institute of Health grants nos. K23 CA92405, CA18029, CA32102, CA114563, R01 CA114563, and R21 CA102624. T.J.B. is the recipient of an American Society of Hematology Junior Faculty Scholar Award.

Authorship

Contribution: S.M. designed the study, contributed to analysis, and wrote manuscript. D.L.S. assisted in data analysis and assisted in writing the manuscript. T.A.A., senior statistician, assisted in

statistical analysis and interpretation of data. T.J.B. provided structural analysis and wrote the paper. R.B.G., statistician, performed statistical analysis. J.L.R. assisted in design of the study. B.J.L., clinical study PI, assisted in data analysis. D.G.G. assisted in design of study. J.P.R. provided valuable resources, assisted in data analysis, and wrote the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Soheil Meshinchi, Fred Hutchinson Cancer Research Center, Clinical Research Division, D5-380, 1100 Fairview Ave N, Seattle, WA 98103; e-mail: smeshinc@fhcrc.org.

References

1. Kiyoi H, Naoe T, Nakano Y, et al. Prognostic implication of FLT3 and N-RAS gene mutations in acute myeloid leukemia. *Blood*. 1999;93:3074-3080.
2. Kottaridis PD, Gale RE, Frew ME, et al. The presence of a FLT3 internal tandem duplication in patients with acute myeloid leukemia (AML) adds important prognostic information to cytogenetic risk group and response to the first cycle of chemotherapy: analysis of 854 patients from the United Kingdom Medical Research Council AML 10 and 12 trials. *Blood*. 2001;98:1752-1759.
3. Thiede C, Steudel C, Mohr B, et al. Analysis of FLT3-activating mutations in 979 patients with acute myelogenous leukemia: association with FAB subtypes and identification of subgroups with poor prognosis. *Blood*. 2002;99:4326-4335.
4. Stirewalt DL, Kopecky KJ, Meshinchi S, et al. FLT3, RAS, and TP53 mutations in elderly patients with acute myeloid leukemia. *Blood*. 2001;97:3589-3595.
5. Meshinchi S, Stirewalt DL, Alonzo TA, et al. Activating mutations of RTK/ras signal transduction pathway in pediatric acute myeloid leukemia. *Blood*. 2003;102:1474-1479.
6. Meshinchi S, Woods WG, Stirewalt DL, et al. Prevalence and prognostic significance of FLT3 internal tandem duplication in pediatric acute myeloid leukemia. *Blood*. 2001;97:89-94.
7. Frohling S, Schlenk RF, Stolze I, et al. Mutations in younger adults with acute myeloid leukemia and normal cytogenetics: prognostic relevance and analysis of cooperating mutations. *J Clin Oncol*. 2004;22:624-623.
8. Meshinchi S, Alonzo TA, Stirewalt DL, et al. Clinical implications of FLT3 mutations in pediatric AML. *Blood*. 2006;108:3654-3661.
9. Stirewalt DL, Kopecky KJ, Meshinchi S, et al. Size of FLT3 internal tandem duplication has prognostic significance in patients with acute myeloid leukemia. *Blood*. 2006;107:3724-3726.
10. Rocnik JL, Okabe R, Yu JC, et al. Roles of tyrosine 589 and 591 in STAT5 activation and transformation mediated by FLT3-ITD. *Blood*. 2006;108:1339-1345.
11. Vempati S, Reindl C, Kaza SK, et al. Arginine 595 is duplicated in patients with acute leukemias carrying internal tandem duplications of FLT3 and modulates its transforming potential. *Blood*. 2007;110:686-694.
12. Lange BJ, Gerbing RB, Feusner J, et al. Mortality in overweight and underweight children with acute myeloid leukemia. *JAMA*. 2005;293:203-211.
13. Zwaan CM, Meshinchi S, Radich JP, et al. FLT3 internal tandem duplication in 234 children with acute myeloid leukemia: prognostic significance and relation to cellular drug resistance. *Blood*. 2003;102:2387-2394.
14. Griffith J, Black J, Faerman C, et al. The structural basis for autoinhibition of FLT3 by the juxtamembrane domain. *Mol Cell*. 2004;13:169-178.
15. Research Collaboratory for Structural Bioinformatics (RCSB), Protein Data Bank. <http://www.rcsb.org/pdb/home/home.do>. Accessed January 15, 2008.
16. Berman HM, Westbrook J, Feng Z, et al. The Protein Data Bank. *Nucleic Acids Res*. 2000;28:235-242.
17. Jones TA, Zou JY, Cowan SW, Kjeldgaard M. Improved methods for building protein models in electron density maps and the location of errors in these models. *Acta Crystallogr A*. 1991;47:110-119.
18. Reindl C, Bagrintseva K, Vempati S, et al. Point mutations in the juxtamembrane domain of FLT3 define a new class of activating mutations in AML. *Blood*. 2006;107:3700-3707.
19. Gale RE, Green C, Allen C, et al. The impact of FLT3 internal tandem duplication mutant level, number, size and interaction with NPM1 mutations in a large cohort of young adult patients with acute myeloid leukemia. *Blood*. 2008;111:2776-2784.
20. Schnittger S, Schoch C, Kern W, Haferlach T. 3' position of the FLT3-LM and loss of heterozygosity are highly correlated to unfavourable outcome in AML. *ASH Annual Meeting Abstracts*. 2006; 108:807.